

Forum Review

Adverse Effects of Reactive Oxygen Species on Vascular Reactivity in Insulin Resistance

DAVID W. BUSIJA, ALLISON W. MILLER, PRASAD KATAKAM, and BENEDEK ERDŐS

ABSTRACT

Insulin resistance (IR) has adverse effects on the reactivity of arteries and arterioles and promotes arterial hypertension and vascular occlusive diseases. Altered reactivity of resistance vessels occurs at both the endothelium and smooth-muscle levels. One major mechanism of vascular dysfunction with IR involves the augmented generation, availability, and/or actions of reactive oxygen species (ROS). Scavengers of ROS are able immediately to restore normal dilator responsiveness in arteries from IR animals. Other factors, such as increased importance of constrictor agents such as endothelin, also restrict normal dilator responses. The basis of ROS-mediated vascular dysfunction in IR may be secondary to underlying inflammatory processes throughout the arterial wall. Although ROS scavengers may be beneficial in the short term, prolonged treatments involving behavioral approaches, such as changes in diet, weight loss, and regular exercise, and pharmacological approaches, involving the use of insulin-sensitizing agents, inhibitors of the renin-angiotensin system, or administration of statins, appear to offer benefits against the detrimental vascular effects of IR. Nonetheless, the most effective approach appears to involve prevention of IR via adoption of a healthy lifestyle by young people. *Antioxid. Redox Signal.* 8, 1131–1140.

INTRODUCTION

INSULIN RESISTANCE (IR) is a major and growing health care problem throughout the world and is a key component of the metabolic syndrome, which represents a major risk factor for the development of cardiovascular disease (1, 14, 19, 21, 30, 33, 37, 38, 42, 46, 92, 104). The reduced ability of insulin to stimulate glucose uptake into skeletal muscle (21) results in glucose intolerance, which is compensated for by prolonged hyperinsulinemia. Eventually, pancreatic β -cells fail to secrete sufficient insulin, leading to non-insulin-dependent diabetes mellitus (NIDDM; type II diabetes). The exact mechanism for the development of IR remains unclear, but current evidence indicates the sequential involvement of obesity-associated visceral adiposity, increased secretion and peripheral actions of adipocytokines, and impairment of insulin-receptor function and/or insulin receptor/intracellular

signal coupling (20, 21, 38, 42, 45, 53, 68, 105). Insulin resistance also is present with many other clinical conditions, such as polycystic ovary syndrome, trauma, and premature birth (10, 11, 18, 56, 64, 65, 79).

The purpose of this review is to summarize our current understanding of the impact of IR on arterial reactivity and to indicate possible underlying mechanisms such as augmented levels of reactive oxygen species (ROS) involved in vascular dysfunction. Although ROS are normal regulators of vascular tone (40), excessive levels of ROS during pathologic conditions appear to lead to inappropriate alterations in reactivity (40, 49). We include information from a variety of arteries to illustrate the diversity of regional circulatory responses that occur in IR. The results discussed are derived largely from studies on rodent models of IR, but experimental data derived from human studies as well as from those of other species are becoming increasingly available and also are discussed.

USEFULNESS OF ANIMAL MODELS OF IR

Examination of the vascular effects of insulin resistance in people is complicated by the presence of other clinical conditions and various medications that affect reactivity. We have found two rat models of IR, the Zucker (fa/fa) obese rat (studied at 12 weeks) and the fructose-fed rat (studied at 10 weeks), particularly useful in our experiments, because we can largely separate the IR syndrome from confounding factors such as hypertension and hyperglycemia. The etiology of IR in the Zucker obese rat is due to a defect in the gene for the leptin receptor (116), whereas a nutritional change leads to IR in the Sprague–Dawley rats on a high-fructose diet (53, 73). During the initial phase of IR, these two types of animals demonstrate severe hyperinsulinemia but have normal fasting blood glucose levels and are not hypertensive (24–26). Fructose-fed rats have normal total cholesterol levels, low high-density lipoprotein levels, and elevated triglyceride levels compared with Sprague–Dawley rats on a normal diet (54, 78). Compared with fructose-fed and normal rats, Zucker obese rats have elevated blood total cholesterol levels and a 3- to 5-times greater elevation of triglyceride levels (32), and therefore the Zucker obese rats can be thought of as having a more advanced or at least a more complicated form of IR. At a later stage of IR in both rat models, vascular dysfunction becomes more severe, and the rats become hypertensive (53). The Zucker obese rats that we use are different from the Zucker diabetic fatty (ZDF) rats, a model of early-onset type II diabetes (119). In addition to the two rat models of IR that we use, other animal models using other strains of rats, mice, pigs, and various other species have been studied and have provided useful information on cardiovascular consequences of IR (85, 117). Although it is more difficult to acquire data on vascular effects of IR from people because of obvious technical and subject-related limitations, it is remarkable how closely the findings from humans parallel those derived from experimental animals.

IMPAIRMENT OF ENDOTHELIUM-DEPENDENT DILATOR RESPONSES

Endothelium produces several important vasodilator substances (32), including prostaglandins, nitric oxide (NO), and endothelium-derived hyperpolarizing factor (EDHF). These substances can diffuse to vascular smooth muscle (VSM), where they activate receptors or second-messenger systems or both, or directly hyperpolarize VSM via gap junctions between endothelium and VSM (32). The relative importance of any one of these endothelium-derived relaxing agents depends on species, age, regional circulation, gender, and vascular segment studied. A consistent finding is that IR reduces the dilator ability of one or more of the endothelium-dependent dilator agents in virtually all of the circulations examined (17, 22, 26, 27, 35, 47, 49, 52, 75, 86, 109). Similar findings concerning impairment of endothelium-dependent dilation in IR people have been derived from several regional circulations (3, 6, 9, 11, 13, 43). Conversely, endothelium-dependent dilator responses can remain intact in arteries from

IR animals (51), and normal responsiveness involving one endothelium-derived relaxing factor may coexist with diminished responsiveness to another factor (86). We are unaware of situations in which dilator responses to any endothelium-derived relaxing factor are enhanced in IR.

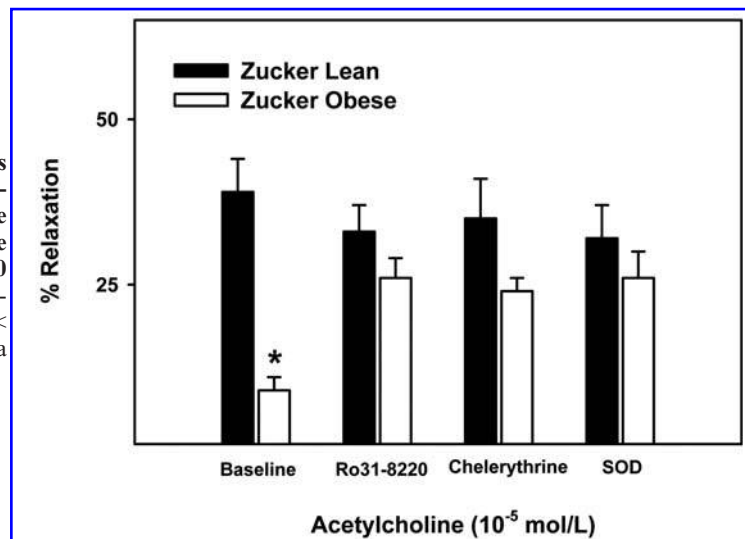
Several factors may lead to reduced endothelium-dependent dilator responsiveness. These include the (a) attenuated production of the dilator substance by endothelium (49, 99–101); (b) augmented degradation of endothelium-derived dilator substances before activating signaling pathways in VSM (26); (c) impaired intracellular signaling in VSM (17, 27, 94, 97, 110, 116); and (d) enhanced production or potency of opposing, endothelium-derived, or neurally-derived constrictor agents (34, 50, 77). Our laboratory has provided examples for each of these situations. First, in the mesenteric arteries of fructose-fed rats, the EDHF component of the dilator responses to bradykinin and acetylcholine is diminished because of a reduction in cytochrome P-450 activity (49). However, the NO-dependent portion to the arterial response to these agents was intact (75). Second, NO-dependent dilation in the basilar artery of Zucker obese rats is impaired because of augmented degradation of NO before it can affect tone of VSM (27) (Fig. 1). The reduced NO-dependent dilator response in the basilar artery is even more striking because levels of endothelial nitric oxide synthase (eNOS) in cerebral arteries are increased in Zucker obese rats (27). Similarly, in coronary arteries of Zucker obese rats, NO-mediated dilation to insulin is reduced in IR despite an increase in eNOS levels (51). Third, prostaglandin-dependent dilation in the middle cerebral artery of fructose-fed rats is reduced because of the impairment of calcium-activated potassium (K_{Ca}) channels coupled to prostaglandin receptors in VSM (25, 26). And fourth, insulin-dependent dilator responses in the mesenteric arteries of fructose-fed rats are reduced in part by enhanced constrictor effects of endothelium-derived endothelin on VSM (77). Enhanced endothelin activity associated with increased receptor expression was observed in mesenteric arteries of fructose-fed rats (50).

IMPAIRMENT OF POTASSIUM CHANNEL FUNCTION IN VSM

The potassium channels involved in mediating responsiveness of arteries and arterioles to various physiologic stimuli arising from the endothelium, blood, perivascular nerves, or parenchyma are the adenosine triphosphate (ATP)-sensitive (K_{ATP}), K_{Ca} , inwardly rectifying (K_{ir}), and voltage-sensitive (K_v) potassium channels (32). Activation of VSM potassium channels with resultant hyperpolarization directly promotes dilation, as in the case of K_{ATP} channels (99), or can act to “buffer” or limit responses to vasoconstrictor stimuli, as occurs with K_{Ca} channels. Additionally, K_{Ca} and K_v channels are tonically active in the cerebral circulation, and inhibition of these channels constrict the cerebral arteries (25–27).

Several studies in experimental animals have shown that VSM potassium channel function is impaired in IR. We are unaware of any studies addressing this issue in people with IR. In the original series of studies in this area, Miller and

FIG. 1. Restoration of normal dilator responses of side branches of the basilar artery to acetylcholine in the Zucker obese rat by protein kinase C inhibitors [Ro31-8220 (5 M)] or chelerythrine (1 μ M)] or superoxide dismutase (SOD; 150 units/ml). Acetylcholine is an endothelium-dependent dilator agent in the basilar artery of the rat. * p < 0.05, compared with response in ZL artery. Data from ref. 27.



colleagues demonstrated that K_{Ca} channel function is impaired in mesenteric arteries of IR rats (17, 49, 52, 73, 74, 76). Impaired K_{Ca} channel function was verified by both *in vitro* diameter and patch-clamp approaches. Conversely, K_{ATP} -mediated dilator responses are intact in this circulation despite IR.

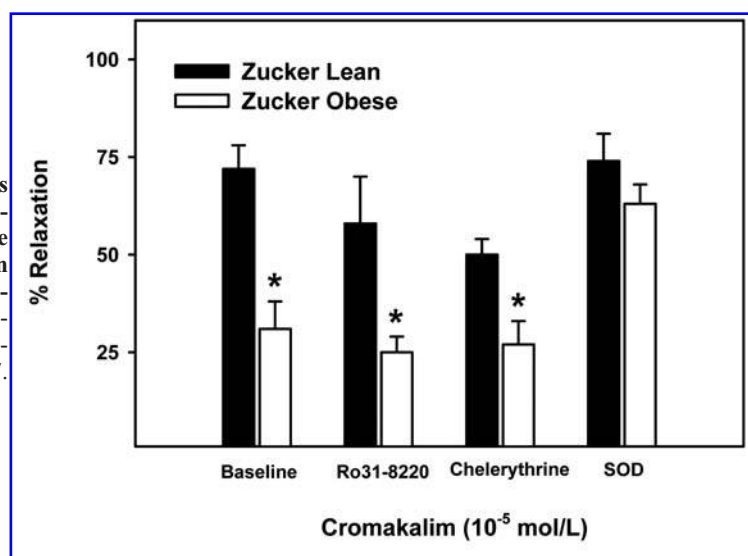
Recently, we showed that both K_{ATP} and K_{Ca} channels are impaired in middle cerebral arteries of fructose-fed IR rats, whereas the K_v and K_{ir} channels are functionally intact (26). Additionally, all four types of potassium channels listed earlier are impaired in the more severely affected Zucker obese rats (27) (Fig. 2). Similar findings were reported recently by another laboratory (86). Impaired potassium channel function is not due to reduced density of K^+ channels in these arteries. By using immunoblot analysis, we found that levels of the $BK_{Ca\alpha}$, K_{ir} 6.1–6.2, and K_{ir} 2.1 proteins (the pore-forming subunits of the BK_{Ca} , K_{ATP} , and K_{ir} channels, respectively) are not detectably affected by IR in either the fructose-fed (26) or Zucker obese (27) rats. These findings are consistent with the

results of a previous study in which the expression of the $BK_{Ca\alpha}$ subunit in mesenteric arteries was found to be unaffected by IR (17).

MECHANISMS OF IMPAIRMENT OF VASCULAR FUNCTION IN IR

The underlying causes of vascular dysfunction in IR are incompletely understood, but we and others have suggested a critical role of ROS resulting from IR-induced vascular inflammation or from other IR-related effects (17, 26, 27, 86, 97) (Figs. 3 and 4). Vascular inflammation, characterized by an elevation in levels of C-reactive protein, interleukin-6, tumor necrosis factor- α , platelet-activating factor, and fibrinogen (1, 12, 19, 42, 91) is associated with elevated ROS levels in arteries from several animal species including humans (27, 41, 48, 99, 101, 115). Augmented vascular ROS levels have been shown to affect en-

FIG. 2. Restoration of normal dilator responses of side branches of the basilar artery to chromakalim in the Zucker obese rat by superoxide dismutase (SOD; 150 units/ml) but not by protein kinase C inhibitors [Ro31-8220 (5 μ M)] or chelerythrine (1 μ M)]. Chromakalim dilates VSM directly by activation of K_{ATP} channels. * p < 0.05, compared with response in ZL artery. Data from ref. 27.



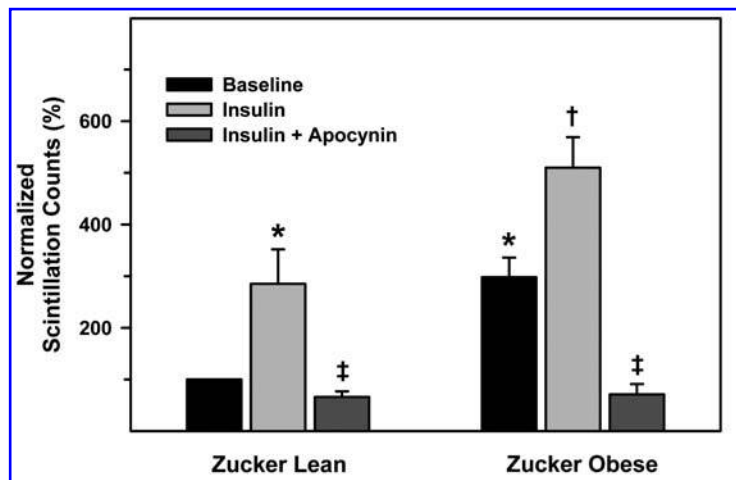


FIG. 3. ROS production in coronary arteries from ZO and ZL rats is higher under baseline conditions and during insulin application (330 ng/ml) and is substantially attenuated by a NAD(P)H oxidase inhibitor (apocynin; 10 μ M). ROS production for the coronary arteries was measured by using the lucigenin-enhanced chemiluminescence method. * $p < 0.05$, compared with ZL baseline. † $p < 0.05$, compared with ZL insulin treatment. ‡ $p < 0.05$, compared with insulin treatment without apocynin. Data from ref. 51.

dothelium- and potassium channel-dependent dilator responses in a variety of settings including IR (2, 4, 26, 27, 62).

ROS could affect vascular reactivity via direct actions on ion channels, receptors, and signaling pathways, or their effects could involve additional steps involving protein kinases and subsequent changes in receptor function or activity of signaling pathways (8, 106). For example, we have shown that ischemia/reperfusion in the cerebral circulation transiently inhibits arterial K_{ATP} function via a ROS-dependent mechanism (4). ROS also are able to degrade NO and thus reduce dilator responses (27), and elevated ROS levels in endothelium may affect levels of enzymes involved in production of dilator agents or inhibit activity of these enzymes (20, 32). Furthermore, impairment of endothelium-dependent dilation in cerebral arteries of Zucker obese rats is reversed by both ROS scavengers and protein kinase C (PKC) inhibitors, thereby suggesting an interaction between ROS and PKC-derived phosphorylation events in promotion of vascular dysfunction

(27) (Figs. 1 and 2). The species of oxygen radicals responsible for IR-mediated dysfunction of cerebral vascular potassium channels and endothelium-dependent response is not known with certainty but, based on recent data from our laboratory, it may be superoxide anion or a superoxide anion-derived radical (26, 27). Nonetheless, identification of the species of oxygen radicals involved is an area deserving further investigation.

Elevated ROS levels are present in various arteries from IR rats in endothelium, VSM, and adventitia (26, 27, 51), which is consistent with the general vascular dysfunction seen in this disease (8, 12, 39, 48, 56, 67, 91, 114) (Fig. 3). A striking finding from our recent studies is that despite established IR in two rat models, application of superoxide dismutase alone or together with catalase leads to immediate restoration of normal responsiveness to endothelium- and potassium channel-dependent dilator stimuli (26, 27) (Figs. 1 and 2). The finding that ROS are able to exert continued inhibitory effects on potassium channel function without causing permanent changes in channel characteristics is supported by patch-clamp studies performed in myocytes from mesenteric arteries of fructose-fed rats (17). Thus, agonist-induced BK_{Ca} -channel activity was impaired when examined in a cell-attached configuration but was normal when the membrane patch was separated from the cell in an inside-out configuration. This finding indicates the presence of endogenous inhibitory substances such as ROS that are able to affect vascular reactivity without permanently changing potassium channel characteristics. However, the duration of IR present in rats and other species is only a matter of a few weeks or several months, and it is unclear whether vascular dysfunction in IR of more prolonged duration, such as decades, as seen in people, would be so readily reversed with ROS scavengers.

Although the underlying etiology and metabolic derangements are different between type I diabetes and IR, subsequent mechanisms of vascular dysfunction appear to be similar for these two diseases. Thus, enhanced ROS levels during type I diabetes also appear to account for reduced endothelium- and VSM potassium channel-dependent dilations in a wide variety of regional circulations from experimental animals and people (31, 40, 119). For example, hyperglycemia-induced production of excessive amounts of superoxide anion enhances degradation of NO (69) and reduces K_v channel

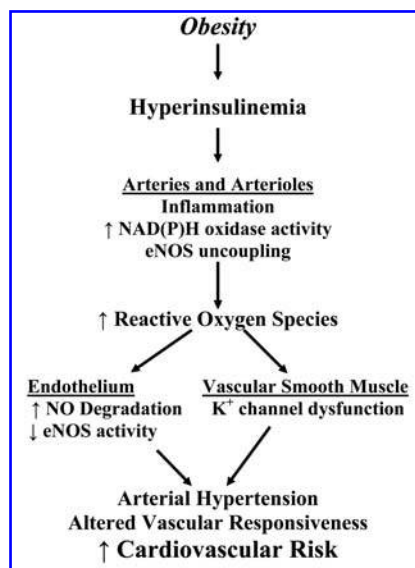


FIG. 4. Schematic depiction of major elements involved in the perturbation of vascular dysfunction in insulin resistance.

opening (63) and K_{ATP} channel function (80) in rat coronary arteries. Additionally, K_{ir} and K_{Ca} channels are impaired in cerebral arteries of diabetic rats (70). Conversely, mechanisms involving hydrogen peroxide but not superoxide anion appear to mediate K_{Ca} channel impairment in type I diabetes (108). Although not so well studied as IR or type I diabetes, ROS also appear to be the mediators of vascular dysfunction in type II diabetes (16, 83, 95).

The augmented ROS production in IR may involve several distinct pathways (2, 7, 41, 59, 81). Recent findings indicate that nicotinamide adenine dinucleotide phosphate, [NAD(P)H]-oxidase activity, an important source of vascular superoxide anion, is augmented in cerebral and coronary arteries from IR rats (26, 27, 107) (Fig. 3). Another source of superoxide anion in IR may arise from the imbalance in levels of eNOS and its essential cofactor, (6R)-5,6,7,8-tetrahydrobiopterin (BH_4). Although eNOS has been shown to be elevated in cerebral and coronary arteries from IR animals (27, 55, 57), perhaps in compensation for reduced NO bioavailability, BH_4 levels are suboptimal, possibly leading to enhanced superoxide anion formation at the expense of NO production because of uncoupling of NOS (41, 99–101). Dietary supplementation of BH_4 leads to decreased oxidative stress and restoration of endothelial function, through increased NO production or NO stability (100). Additional pathways leading to increased ROS production during IR may involve metabolism of arachidonic acid via several enzymatic pathways (2), metabolism of xanthine by xanthine oxidase system (32), or enhanced production of superoxide anion from mitochondria (36).

Whereas ROS appear to be a major cause of reduced dilator responses in IR, other mechanisms such as enhanced constrictor effects via increased sympathetic nervous system activity (34) or augmented release of endothelin (77) also may be involved in some situations. It is unclear whether an increased contribution to vascular tone by sympathetic nerves or endothelin is due to specific effects of hyperinsulinemia or is secondary to general inflammatory responses associated with IR or both. Increases in responses to constrictor agents during IR apparently do not occur in all regional circulations. We have found that constrictor effects in the middle cerebral artery (28) or basilar artery (103) are unaffected in IR, but constrictor responses are enhanced in mesenteric arteries (77).

STRATEGIES TO RESTORE NORMAL VASCULAR FUNCTION IN IR

A variety of dietary, behavioral, and pharmacologic interventions have been directed toward improving vascular function by reversing or lessening the severity of IR, by delaying the progression to NIDDM, and by directly targeting the vasculature to restore normal responsiveness. Nonpharmacologic approaches involve weight loss (6, 38, 44, 66, 72, 102, 115), consumption of a balanced diet rich in antioxidants (38, 44, 66, 72, 102), and regular exercise (6, 38, 60, 72). Although these approaches have shown some benefit against IR in people and in animal models, it is unclear whether these interventions alone will lead to sustained restoration of normal

responsiveness of the vasculature (6), especially with long-standing IR, and therefore more research is needed for alternative therapies (6, 37, 44, 112). Another concern is whether these approaches can be sustained for extended periods in aging individuals with other conditions that may interfere with maintaining regular physical activity or adhering to a special diet.

Several pharmacologic agents, which offer the benefits of rapid improvement, convenience, and sustained benefit, have been shown to be effective in improving vascular function in IR. For example, administration of ROS scavengers, such as superoxide dismutase and catalase, is able to restore normal vascular responsiveness by endothelium and VSM almost immediately in rodent models of IR (17, 26, 27) (Figs. 1 and 2). It also has been shown that increased dietary intake of appropriate antioxidants is an effective treatment in people against a variety of vascular-related problems (43, 87, 111). General appreciation exists of the health-related value of increasing the body level of diet-derived antioxidants (43, 87, 111). These agents are readily available in natural foods and over-the-counter supplements, and especially the water-soluble ones have few significant side effects, even with prolonged consumption. Furthermore, oral administration of the eNOS cofactor, BH_4 , also restores endothelium function in an animal model of IR, probably by favoring NO production rather than superoxide anion formation (100, 101). Endothelium-dependent dilator effects also are improved with topical application of PKC inhibitors in IR rats, although IR-dependent suppression of VSM potassium channel function is not restored (27) (Figs. 1 and 2). However, the relation between PKC activation, ROS availability, and vascular reactivity in IR is complex (2, 61, 84, 88, 106). It may not be practical or beneficial to administer agents to augment BH_4 levels or to block activation of PKC for long periods because of unproven effectiveness and specificity.

Other promising pharmacologic approaches currently in widespread clinical use involve administration of insulin-sensitizing drugs, statins, or drugs acting against the renin-angiotensin system. Metformin and thiazolidinediones have been shown to improve endothelial function in experimental animals and people and to restore NO-dependent dilation (15, 23, 54, 71, 73, 82, 112), probably by promoting improvement in the IR condition (112). Statins are effective in restoring vascular function in IR in rats (18, 22, 29, 45, 78, 112) perhaps by reducing vascular inflammation and decreasing vascular levels of ROS (18, 29, 38, 78) or promoting expression of enzymes involved in the synthesis of dilator stimuli (18, 22) rather than by reducing cholesterol levels in blood. We have shown that treatment with a clinical dose of a statin in fructose-fed or Zucker obese rats is able to restore coronary and cerebral artery function (29, 78). Another pharmacologic approach that has been shown to be beneficial, especially in diabetes, involves targeting the renin-angiotensin system. Obesity, which is a major risk factor for the development of IR, leads to increased sympathetic nervous system activity and production of angiotensin II-forming enzymes and subsequent arterial hypertension and organ damage (8, 89, 98). Administration of angiotensin-converting enzyme inhibitors and angiotensin type-1 receptor blockers have been shown to reduce arterial hypertension and also to protect the heart and kidneys (8, 89, 93, 98).

An insidious and largely unappreciated aspect of IR is that detrimental effects on the cardiovascular system are taking place for a substantial period before the detection of the disease. Traditional diagnostic factors, such as fasting blood glucose levels and arterial blood pressure, may be within normal limits during much of the progression of IR. Impairment of vascular function during this period, which may interfere with coupling between metabolism and blood flow, may lead to reduced exercise tolerance and cognitive function. Insulin resistance appears to be a risk factor for Alzheimer disease (39, 58, 67, 90, 96, 113), and chronic IR has been associated with organ damage in people (14, 19, 94). Furthermore, the combination of arterial hypertension with IR or NIDDM represents an especially serious challenge to the cardiovascular system and will greatly increase the risk of atherosclerosis, coronary occlusive disease, and strokes in these patients. Therefore, young people with risk factors for IR, such as family history and obesity, should likely be more aggressively tested and treated for IR before the development of long-term complications.

PERSPECTIVES

Insulin resistance leads to impairment of normal vascular function and therefore is a major risk factor for development of cardiovascular disease. Vascular dysfunction caused by IR, at least in experimental animals, involves effects of augmented ROS produced within the vessel wall through a number of possible metabolic pathways including NAD(P)H oxidase. More research is needed in defining the specific mechanisms of impairment, especially with respect to potassium channels in VSM. Although reduced endothelium and VSM effects are reversed by ROS scavengers in animals after weeks or months of IR, it is not clear whether vascular dysfunction associated with prolonged IR, as occurs in people, would be so easily remedied. Behavioral, dietary, and/or pharmacologic approaches that prevent or reverse the underlying inflammatory process have great potential for minimizing vascular impairment and subsequent development of cardiovascular diseases associated with IR. However, additional research must be done in defining strategies for treatment in patients with long-standing IR or NIDDM.

ACKNOWLEDGMENTS

This study was supported by grants HL-30260, HL-50587, HL-66074, HL-65380, HL-77731, DK-62372 from NIH, and a Bugher Foundation Award through the American Heart Association (0270114N). B.E. was supported by a Hungarian National Eötvös scholarship.

ABBREVIATIONS

K_{ATP} channel, ATP-sensitive potassium channel; EDHF, endothelium-derived hyperpolarizing factor; K_{Ca} channel, calcium-activated potassium channel; eNOS, endothelial ni-

tric oxide synthase; K_{ir} channel, inwardly rectifying potassium channel; IR, insulin resistance; NAD(P)H oxidase, nicotinamide adenine dinucleotide phosphate oxidase; NIDDM, non-insulin-dependent diabetes mellitus; NO, nitric oxide; PKC, protein kinase C; ROS, reactive oxygen species; BH₄, (6R)-5,6,7,8-tetrahydrobiopterin; VSM, vascular smooth muscle; K_v potassium channels, voltage-sensitive potassium channel; ZDF rats, Zucker diabetic fatty rats; ZL rats, Zucker lean rats; ZO rats, Zucker obese rats.

REFERENCES

- Abbatecola AM, Ferrucci L, Grella R, Bandinelli S, Bonafe M, Barbieri Corsi AM, Lauretani F, Franceschi C, and Paolisso G. Diverse effect of inflammatory markers on insulin resistance and insulin resistant syndrome in the elderly. *J Am Geriatr Soc* 52: 399–404, 2004.
- Armstead WM. Vasopressin-induced protein kinase C-dependent superoxide generation contributes to ATP-sensitive potassium channel but not calcium-sensitive potassium channel function impairment after brain injury. *Stroke* 32: 1408–1414, 2001.
- Balletshofer BM, Rittig K, Stock J, Lehn-Stefan A, Overkamp D, Dietz K, and Haring HU. Insulin resistant young subjects at risk of accelerated atherosclerosis exhibit a marked reduction in peripheral endothelial function early in life but not differences in intima-media thickness. *Atherosclerosis* 171: 303–309, 2003.
- Bari F, Louis TM, Meng W, and Busija DW. Global ischemia impairs ATP-sensitive K⁺ channel function in cerebral arterioles in piglets. *Stroke* 27: 1874–1881, 1996.
- Bloomgarden ZT. Mediators, pediatric insulin resistance, the polycystic ovary syndrome, and malignancy. *Diabetes Care* 28: 1821–1830, 2005.
- Brook RD, Bard RL, Glazewski L, Kehrer C, Bodary PF, Eitzman DL, Rajagopalan S. Effect of short-term weight loss on the metabolic syndrome and conduit vascular endothelial function in overweight adults. *Am J Cardiol* 93: 1012–1016, 2004.
- Busija DW. Prostaglandins and other eicosanoids. In: *Cerebral Blood Flow and Metabolism*, Edvinsson L and Krause D, eds. Philadelphia: Lippincott Williams & Wilkins, 2002; 325–338.
- Ceriello A and Motz E. Is oxidative stress the pathogenic mechanism underlying insulin resistance, diabetes, and cardiovascular disease? The common soil hypothesis revisited. *Arterioscler Thromb Vasc Biol* 24: 816–823, 2004.
- Campia U, Sullivan G, Bryant MB, Waclawiw MA, Quon MJ, and Panza JA. Insulin impairs endothelium-dependent vasodilation independent of insulin sensitivity or lipid profile. *Am J Physiol Heart Circ Physiol* 286: H76–H82, 2004.
- Carlson GL. Hunterian Lecture: Insulin resistance in human sepsis: Implications for the nutritional and metabolic care of the critically ill surgical patient. *Ann R Coll Surg Engl* 86: 75–81, 2004.
- Carmassi F, De Negri F, Fioriti R, DeGiorgi A, Giannarelli C, Fruzzetti F, Pedrinelli R, Dell'omo G, and Bersi C. In-

- sulin resistance causes impaired vasodilation and hypofibrinolysis in young women with polycystic ovary syndrome. *Thromb Res* 116: 207–214, 2005.
12. Conti M, Renaud IM, Poirier B, Michel O, Belair MF, Mandet C, Brune P, Myara I, and Chevalier J. High levels of myocardial antioxidant defense in aging nondiabetic normotensive Zucker obese rats. *Am J Physiol Regul Integr Comp Physiol* 286: R793–R800, 2004.
 13. Dagres N, Saller B, Haude M, Husing J, von Birgelen C, Schmermund A, Sack S, Baumgart D, Mann K, and Erbel R. Insulin sensitivity and coronary vasoreactivity: Insulin sensitivity relates to adenosine-stimulated coronary flow response in human subjects. *Clin Endocrinol* 61: 724–731, 2004.
 14. de la Torre JC. Impaired cerebrovascular perfusion: summary of evidence in support of its causality in Alzheimer's disease. *Ann NY Acad Sci* 924: 136–152, 2000.
 15. Diamanti-Kandarakis E, Alexandraki K, Protogerou A, Piperi C, Papamichael C, Aessopos A, Lekakis J, and Mavrikakis M. Metformin administration improves endothelial function in women with polycystic ovary syndrome. *Eur J Endocrinol* 152: 749–756, 2005.
 16. Didion SP, Lynch CM, Baumbach GL, and Faraci FM. Impaired endothelium-dependent responses and enhanced influences of Rho-kinase in cerebral arterioles in type II diabetes. *Stroke* 36: 342–347, 2005.
 17. Dimitropoulou C, Han G, Miller AW, Molero M, Fuchs LC, White RE, and Carrier GO. Potassium (BK(Ca)) currents are reduced in microvascular smooth muscle cells from insulin-resistant rats. *Am J Physiol Heart Circ Physiol* 282: H908–H917, 2002.
 18. Di Napoli M. Benefits of statins in cerebrovascular disease. *Invest Drugs* 5: 295–305, 2004.
 19. Dogra GK, Hermann S, Irish AB, Thomas MA, and Watts GF. Insulin resistance, dyslipidaemia, inflammation and endothelial function in nephritic syndrome. *Nephrol Dial Transplant* 17: 2220–2225, 2002.
 20. Domoki F, Velkamp R, Thrikawala N, Robins G, Bari F, Louis TM, and Busija DW. Ischemia-reperfusion rapidly increases COX-2 expression in piglet cerebral arteries. *Am J Physiol Heart Circ Physiol* 277: H1207–H1214, 1999.
 21. Donnelly R and Qu X. Mechanisms of insulin resistance and new pharmacologic approaches to metabolism and diabetic complications. *Clin Exp Pharmacol Physiol* 25: 79–87, 1998.
 22. Dumont AS, Hyndman ME, Dumont RJ, Fedak PM, Kasell NF, Sutherland GR, and Verma S. Improvement of endothelial function in insulin-resistant carotid arteries treated with pravastatin. *J Neurosurg* 95: 466–471, 2001.
 23. Durbin RJ. Thiazolidinedione therapy in the prevention/delay of type 2 diabetes in patients with impaired glucose tolerance and insulin resistance. *Diabetes Obesity Metab* 6: 280–285, 2004.
 24. Erdős B, Miller AW, and Busija DW. Impaired endothelium-mediated relaxation in isolated cerebral arteries from insulin-resistant rats. *Am J Physiol Heart Circ Physiol* 282: H2060–H2065, 2002.
 25. Erdős B, Miller AW, and Busija DW. Alterations in K_{ATP} and K_{Ca} channel function in cerebral arteries of insulin resistant rats. *Am J Physiol Heart Circ Physiol* 283: H2472–H2477, 2002.
 26. Erdős B, Simandle SA, Snipes JA, Miller AW, and Busija DW. Potassium channel dysfunction in cerebral arteries of insulin-resistant rats is mediated by reactive oxygen species. *Stroke* 35: 964–969, 2004.
 27. Erdős B, Snipes JA, Miller AW, and Busija DW. Cerebrovascular dysfunction in Zucker obese rats is mediated by oxidative stress and protein kinase C. *Diabetes* 53: 1352–1359, 2004.
 28. Erdős B, Snipes JA, Kis B, Miller AW, and Busija DW. Vasoconstrictor mechanisms in the cerebral circulation are unaffected by insulin resistance. *Am J Physiol Regul Integr Comp Physiol* 287: R1456–R1461, 2004.
 29. Erdős B, Snipes JA, Tulbert CD, Katakam P, Miller AW, and Busija DW. Rosuvastatin improves cerebrovascular function in Zucker obese rats by inhibiting NAD(P)H-oxidase-dependent superoxide production. *Am J Physiol Heart Circ Physiol* 290(3): H1264–1270, 2006.
 30. Eschwege E. The dysmetabolic syndrome, insulin resistance and increased cardiovascular (CV) morbidity and mortality in type 2 diabetes: Aetiological factors in the development of CV complications. *Diabetes Metab* 29: 6S19–6S27, 2003.
 31. Faraci FM. Oxidative stress: The curse that underlies cerebral vascular dysfunction? *Stroke* 36: 186–188, 2005.
 32. Faraci FM and Heistad DD. Regulation of potassium channels in regulation of cerebral circulation: Role of endothelium and potassium channels. *Physiol Rev* 78: 53–97, 1998.
 33. Ford ES, Giles WH, and Dietz WH. Prevalence of the metabolic syndrome among U.S. adults: Findings from the third National Health and Nutrition Examination Survey. *J Clin Invest* 287: 356–359, 2002.
 34. Frisbee JC. Enhanced arteriolar α -adrenergic constriction impairs dilator responses and skeletal muscle perfusion in obese Zucker rats. *J Appl Physiol* 97: 764–772, 2004.
 35. Frisbee JC and Stepp DW. Impaired NO-dependent dilation of skeletal muscle arterioles in hypertensive diabetic obese Zucker rats. *Am J Physiol Heart Circ Physiol* 281: H1304–1311, 2001.
 36. Garlid KD and Paucek P. Mitochondrial potassium transport: the K (+) cycle. *Biochim Biophys Acta* 1606: 23–41, 2003.
 37. Grant RW and Meigs JB. Should the insulin resistance syndrome be treated in the elderly? *Drugs Aging* 21: 141–151, 2004.
 38. Grundy SM, Hansen B, Smith SC, Cleeman JI, and Kahn RA, for Conference Participants. Clinical management of metabolic syndrome: Report of the American Heart Association/National Heart, Lung, and Blood Institute/American Diabetes Association conference on scientific issues related to management. *Circulation* 109: 551–556, 2004.
 39. Gustafson D, Rothenberg E, Blennow K, Steen B, and Skoog I. An 18-year follow-up of overweight and risk of Alzheimer disease. *Arch Intern Med* 163: 1524–1528, 2003.
 40. Gutterman DD, Miura H, and Liu Y. Redox modulation of vascular tone: Focus of potassium channel mechanisms of dilation. *Arterioscler Thromb Vasc Biol* 25: 671–678, 2005.
 41. Guzik TJ, Mussa S, Gastaldi D, Sadowski J, Ratnatunga C, Pillai R, and Channon KM. Mechanisms of increased vas-

- cular superoxide production in human diabetes mellitus: Role of NAD(P)H oxidase and endothelial nitric oxide synthase. *Circulation* 105: 1656–1662, 2002.
42. Hidvegi T, Szatmari F, Hetyesi K, Biro L, and Jermendy G. Intima-media thickness of the carotid arteries in subjects with hyperinsulinaemia (insulin resistance). *Diabetes Nutr Metab* 16: 139–144, 2003.
 43. Hirashima O, Kawano H, Motoyama T, Hirai N, Ohgushi M, Kugiyama K, Ogawa H, and Yasue H. Improvement of endothelial function and insulin sensitivity with vitamin C in patients with coronary spastic angina: possible role of reactive oxygen species. *J Am Coll Cardiol* 35: 1860–1866, 2000.
 44. Holness MJ, Smith ND, Greenwood GK, and Sugden MC. Acute omega-3 fatty acid enrichment selectively reverses high-saturated fat feeding-induced insulin hypersecretion but does not improve peripheral insulin resistance. *Diabetes* 53(suppl 1): S166–S171, 2004.
 45. Hotamisligil GS. The role of TNF α and TNF receptors in obesity and insulin resistance. *J Intern Med* 245: 621–625, 1999.
 46. Jorgensen H, Nakayama H, Raaschou HO, and Olsen TS. Stroke in patients with diabetes: The Copenhagen Stroke Study. *Stroke* 25: 1977–1984, 1994.
 47. Karagiannis J, Reid JJ, Darby I, Roche P, Rand MJ, and Li CG. Impaired nitric oxide function in the basilar artery of the obese Zucker rat. *J Cardiovasc Pharmacol* 42: 497–505, 2003.
 48. Kashiwagi A, Shinozaki K, Nishio Y, Okamura T, Toda N, and Kikkawa R. Free radical production in endothelial cells as a pathogenetic factor for vascular dysfunction in the insulin resistance state. *Diabetes Res Clin Pract* 45: 199–203, 1999.
 49. Katakam PV, Hoenig M, Ujhelyi MR, and Miller AW. Cytochrome P450 activity and endothelial dysfunction in insulin resistance. *J Vasc Res* 37: 426–434, 2000.
 50. Katakam PV, Pollock JS, Pollock DM, Ujhelyi MR, and Miller AW. Enhanced endothelin-1 response and receptor expression in small mesenteric arteries of insulin-resistant rats. *Am J Physiol Heart Circ Physiol* 280: H522–H527, 2001.
 51. Katakam PV, Tulbert CD, Snipes JA, Erdos B, Miller AW, and Busija DW. Impaired insulin-induced vasodilation in small coronary arteries of Zucker obese rats is mediated by reactive oxygen species. *Am J Physiol Heart Circ Physiol* 288: H854–H860, 2005.
 52. Katakam PV, Ujhelyi MR, and Miller AW. EDHF-mediated relaxation is impaired in fructose-fed rats. *J Cardiovasc Pharmacol* 34: 461–467, 1999.
 53. Katakam PV, Ujhelyi MR, Hoenig ME, and Miller AW. Endothelial dysfunction precedes hypertension in diet-induced insulin resistance. *Am J Physiol* 275: R788–R792, 1998.
 54. Katakam PV, Ujhelyi MR, Hoenig M, and Miller A.W. Metformin improves vascular function in insulin-resistant rats. *Hypertension* 35: 108–112, 2000.
 55. Kawaguchi M, Koshimura K, Sohmiya M, Murakami Y, Gonda T, and Kato Y. Effect of insulin on nitric oxide synthase-like immunostaining of arteries in various organs in Zucker diabetic fatty rats. *Eur J Endocrinol* 145: 343–349, 2001.
 56. Kelly CJ, Speirs A, Gould GW, Petrie JR, Lyall H, and Connell JM. Altered vascular function in young women with polycystic ovary syndrome. *J Clin Endocrinol Metab* 87: 742–746, 2002.
 57. Kuboki K, Jiang ZY, Takahara N, Ha SW, Igarashi M, Yamauchi T, Feener EP, Herbert TP, Rhodes CJ, and King GL. Regulation of endothelial constitutive nitric oxide synthase gene expression in endothelial cells and in vivo: A specific vascular action of insulin. *Circulation* 101: 676–681, 2000.
 58. Kudo T, Imaizumi K, Tanimukai H, Katayama T, Sato N, Nakamura Y, Tanaka T, Kashiwagi Y, Jinno Y, Tohyama M, and Takeda M. Are cerebrovascular factors involved in Alzheimer's disease? *Neurobiol Aging* 21: 215–224, 2000.
 59. Lacza Z, Puskar M, Kis B, Perciaccante JV, Miller AW, and Busija DW. Hydrogen peroxide acts as an EDHF in the piglet pial vasculature in response to bradykinin. *Am J Physiol Heart Circ Physiol* 281: H406–H411, 2002.
 60. Lamonte MJ, Blair SN, and Church TS. Physical activity and diabetes prevention. *J Appl Physiol* 99: 1205–1213, 2005.
 61. Lee HB, Yu MR, Song JS, and Ha H. Reactive oxygen species amplify protein kinase C signaling in high glucose-induced fibronectin expression by human peritoneal mesothelial cells. *Kidney Int* 65: 1170–1179, 2004.
 62. Liu Y and Gutterman DD. Oxidative stress and potassium channel function. *Clin Exp Pharmacol Physiol* 29: 305–311, 2002.
 63. Liu Y, Terata K, Rusch NJ, and Gutterman DD. High glucose impairs voltage-gated K(+) channel current in rat small coronary arteries. *Circ Res* 89: 146–152, 2001.
 64. Ma Y, Toth B, Keeton AB, Holland LT, Chaudry IH, and Messina JL. Mechanisms of hemorrhage-induced hepatic insulin resistance: role of tumor necrosis factor- α . *Endocrinology* 145: 5168–5176, 2004.
 65. Ma Y, Wang P, Kuebler JF, Chaudry IH, and Messina JL. Hemorrhage induces the rapid development of hepatic insulin resistance. *Am J Physiol Gastrointest Liver Physiol* 284: G107–G115, 2003.
 66. Maki KC. Dietary factors in the prevention of diabetes mellitus and coronary artery disease associated with the metabolic syndrome. *Am J Cardiol* 93(suppl): 12C–17C, 2004.
 67. Matsumoto K, Miyake S, Yano M, Ueki Y, Miyazaki A, Hirao K, and Tominaga Y. Insulin resistance and classic risk factors in type 2 diabetic patients with different subtypes of ischemic stroke. *Diabetes Care* 22: 1191–1195, 1999.
 68. Matsuzawa Y, Funahashi T, Nakamura T. Molecular mechanism of metabolic syndrome X: Contribution of adipocytokines adipocyte-derived bioactive substances. *Ann NY Acad Sci* 892: 146–154, 1999.
 69. Mayhan WG. Superoxide dismutase partially restores impaired dilatation of the basilar artery during diabetes mellitus. *Brain Res* 760: 204–209, 1997.
 70. Mayhan WG, Mayhan JF, Sun H, and Patel KP. In vivo properties of potassium channels in cerebral blood vessels during diabetes mellitus. *Microcirculation* 11: 605–613, 2004.
 71. Meriden T. Progress with thiazolidinediones in the management of type 2 diabetes mellitus. *Clin Ther* 26: 177–190, 2004.

72. Metzger BL. The effect of a genetic variant for obesity and type 2 diabetes on the therapeutic potential of exercise and caloric restrictive diets in Zucker rats. *Res Theory Nurs Pract* 17: 321–333, 2003.
73. Miatello R, Cruzado M, and Risler N. Mechanisms of cardiovascular changes in an experimental model of syndrome X and pharmacological intervention on the renin-angiotensin system. *Curr Vasc Pharmacol* 2: 371–377, 2004.
74. Miller AW, Dimitropoulou C, Han G, White RE, Busija DW, and Carrier GO. Epoxyeicosatrienoic acid-induced relaxation is impaired in insulin resistance. *Am J Physiol Heart Circ Physiol* 281: H1524–H1531, 2001.
75. Miller AW, Hoenig ME, and Ujhelyi MR. Mechanisms of impaired endothelial function associated with insulin resistance. *J Cardiovasc Pharmacol Ther* 3: 125–134, 1998.
76. Miller AW, Katakam PV, and Ujhelyi MR. Impaired endothelium-mediated relaxation in coronary arteries from insulin-resistant rats. *J Vasc Res* 36: 385–392, 1999.
77. Miller AW, Tulbert CD, Puskar M, and Busija DW. Enhanced endothelin activity prevents vasodilation to insulin in insulin resistance. *Hypertension* 40: 78–82, 2002.
78. Miller AW, Tulbert CD, and Busija DW. Rosuvastatin treatment reverses impaired coronary artery vasodilation in fructose-fed, insulin resistant rats. *Am J Physiol Regul Integr Comp Physiol* 287: R157–R160, 2004.
79. Mitanchez-Mokhtari D, Lahlou N, Kieffer F, Magny JF, Roger M, and Voyer M. Both relative insulin resistance and defective islet beta-cell processing of proinsulin are responsible for transient hyperglycemia in extremely preterm infants. *Pediatrics* 113: 537–541, 2004.
80. Miura H, Wachtel RE, Loberiza FR Jr, Saito T, Miura M, Nicolosi AC, and Gutterman DD. Diabetes mellitus impairs vasodilation to hypoxia in human coronary arterioles: Reduced activity of ATP-sensitive potassium channels. *Circ Res* 92: 151–158, 2003.
81. Mohazzab KM, Kaminski PM, and Wolin MS. NADH oxidoreductase is a major source of superoxide anion in bovine coronary artery endothelium. *Am J Physiol Heart Circ Physiol* 266: H2586–H2572, 1994.
82. Natali A, Baldeweg S, Toschi E, Capaldo B, Barbaro D, Gastaldelli A, Yudkin JS, and Ferrannini E. Vascular effects of improving metabolic control with metformin or rosiglitazone in type 2 diabetes. *Diabetes Care* 27: 1349–1357, 2004.
83. Okon EB, Chug AW, Rauniyar P, Padilla E, Tejerina T, McManus BM, Luo H, and van Breemen C. Compromised arterial function in human type 2 diabetic patients. *Diabetes* 54: 2415–2423, 2005.
84. Otani H. Reactive oxygen species as mediators of signal transduction in ischemic preconditioning. *Antioxid Redox Signal* 6: 449–469, 2004.
85. Pamies-Andreu E, Fiksen-Olsen M, Rizza RA, Romero JC. High-fructose feeding elicits insulin resistance without hypertension in normal mongrel dogs. *Am J Hypertens* 8: 732–738, 1995.
86. Phillips SA, Sylvester FA, and Frisbee JC. Oxidant stress and constrictor reactivity impair cerebral artery dilation in obese Zucker rats. *Am J Physiol Regul Integr Comp Physiol* 288: R522–R530, 2005.
87. Pleiner J, Schaller G, mittermayer F, Bayerle-Eder M, Roden M, and Wolzt M. FFA-induced endothelial dysfunction can be corrected by vitamin C. *J Clin Endocrinol Metab* 87: 2913–2917, 2002.
88. Pricci F, Leto G, Amadio L, Iacobini C, Cordone S, Catalano S, Zicari A, Sorcini M, Di Mario U, and Pugliese G. Oxidative stress in diabetes-induced endothelial dysfunction: involvement of nitric oxide and protein kinase C. *Free Radic Biol Med* 35: 683–694, 2003.
89. Quinones MJ, Hernandez-Pamploni M, Schelbert H, Bulnes-Enriquez, Jimenez X, Hernandez G, De La Rosa R, Chon Y, Yang H, Nicholas SB, Modilevsky T, Yu K, Van Herle K, Castellani LW, Elashoff R, and Hsueh WA. Coronary vasomotor abnormalities in insulin-resistant individuals. *Ann Intern Med* 140: 700–708, 2004.
90. Razay G and Wilcock GK. Hyperinsulinaemia and Alzheimer's disease. *Age Ageing* 23: 396–399, 1994.
91. Sakkinen PA, Whal P, Cushman M, Lewis MR, and Tracy RP. Clustering of procoagulation, inflammation, and fibrinolysis variables with metabolic factors in insulin resistance syndrome. *Am J Epidemiol* 152: 897–907, 2000.
92. Salonen JT, Lakka TA, Lakka HM, Valkonen VP, Everson SA, and Kaplan GA. Hyperinsulinemia is associated with the incidence of hypertension and dyslipidemia in middle-aged men. *Diabetes* 47: 270–275, 1998.
93. Scheen AJ. Renin-angiotensin system inhibition prevents type 2 diabetes mellitus: Part 2: Overview of physiological and biochemical mechanism. *Diabetes Metab* 30: 498–505, 2004.
94. Schunkert H. Obesity and target organ damage: The heart. *Int J Obes Relat Metab Disord* 4: S15–S20, 2002.
95. Schwaninger RM, Sun H, and Mayhan WG. Impaired nitric oxide synthase-dependent dilatation of cerebral arterioles in type II diabetic rats. *Life Sci* 73: 3415–3425, 2003.
96. Shinozaki K, Naritomi H, Shimizu T, Suzuki M, Ikebuchi M, Sawada T, and Harano Y. Role of insulin resistance associated with compensatory hyperinsulinemia in ischemic stroke. *Stroke* 27: 37–43, 1996.
97. Sjöholm A and Nystrom T. Endothelial inflammation in insulin resistance. *Lancet* 365: 610–612, 2005.
98. Sharma AM. Is there a rationale for angiotensin blockade in the management of obesity hypertension? *Hypertension* 44: 12–19, 2004.
99. Shinozaki K, Hirayama A, Nishio Y, Yoshida Y, Ohtani T, Okamura T, Masada M, Kikkawa R, Kodama K, and Kashiwagi A. Coronary endothelial dysfunction in the insulin-resistant state is linked to abnormal pteridine metabolism and vascular oxidative stress. *J Am Coll Cardiol* 38: 1821–1828, 2001.
100. Shinozaki K, Kashiwagi A, Masada M, and Okamura T. Molecular mechanisms of impaired endothelial function associated with insulin resistance. *Curr Drug Targets Cardiovasc Haematol Disord* 4: 1–11, 2004.
101. Shinozaki K, Kashiwagi A, Nishio Y, Okamura T, Yoshida Y, Masada M, Toda N, and Kikkawa R. Abnormal biopterin metabolism is a major cause of impaired endothelium-dependent relaxation through nitric oxide/ O_2^- imbalance in insulin-resistant rat aorta. *Diabetes* 48: 2437–2445, 1999.

102. Shirai K. Obesity as the core of the metabolic syndrome and the management of coronary heart disease. *Curr Med Res Opin* 20: 295–304, 2004.
103. Simandle SA, Erdős B, Snipes JA, Miller AW, and Busija DW. Insulin resistance does not impair contractile responses of cerebral arteries. *Life Sci* 77: 2262–2272, 2005.
104. Steinberg HO and Baron AD. Vascular function, insulin resistance and fatty acids. *Diabetologia* 45: 623–634, 2002.
105. Stepan CM and Lazar MA. Resistin and obesity-associated insulin resistance. *Trends Endocrinol Metab* 13: 18–23, 2002.
106. Sowers JR. Insulin resistance and hypertension. *Am J Physiol Heart Circ Physiol* 286: H1597–H1602, 2004.
107. Sun CK, Zhang XY, and Wheatley AM. Increased NAD(P)H fluorescence with decreased blood flow in the steatotic liver of the obese Zucker rat. *Microvasc Res* 66: 15–21, 2003.
108. Tang XD, Garcia ML, Heinemann SH, and Hoshi T. Reactive oxygen species impairs Slo1 BK channel function by altering cysteine-mediated calcium sensing. *Nat Struct Mol Biol* 11: 171–178, 2004.
109. Turner R (UK Prospective Diabetes Study UKPDS Group). Effect of intensive blood-glucose control with metformin on complications in overweight patients with type 2 diabetes (UKPDS 34). *Lancet* 352:854–865, 1998.
110. Turtle JR. The economic burden of insulin resistance. *Int J Clin Pract Suppl* 113: 23–28, 2000.
111. Voko Z, Hollander M, Hofman A, Koudstaal PJ, and Breteler MM. Dietary antioxidants and the risk of ischemic stroke: The Rotterdam study. *Neurology* 61: 1273–1275, 2003.
112. Wagh A and Stone NJ. Expert review of cardiovascular therapy. *Exp Rev Cardiovasc Ther* 2: 213–228, 2004.
113. Watson GS and Craft S. The role of insulin resistance in the pathogenesis of Alzheimer's disease: Implications for treatment. *CNS Drugs* 17: 27–45, 2003.
114. Wiernsperger NF and Bouskela E. Microcirculation in insulin resistance and diabetes: more than just a complication. *Diabetes Metab* 29: 6S77–6S87, 2003.
115. Wisse BE. The inflammatory syndrome: The role of adipose tissue cytokines in metabolic disorders linked to obesity. *J Am Soc Nephrol* 15: 2792–2800, 2004.
116. Yamashita T, Murakami T, Iida M, Kuwajima M, and Shima K. Leptin receptor of Zucker fatty rat performs reduced signal transduction. *Diabetes* 46: 1077–1080, 1997.
117. Vollenwieder P. Insulin resistant states and insulin signaling. *Clin Chem Lab Med* 41: 1107–1119, 2003.
118. Xiang AH, Peters RK, Kjos SL, Goico J, Ochoa C, Marroquin A, Tan S, Hodis HN, Azen SP, and Buchanan TA. Pharmacological treatment of insulin resistance at two different stages in the evolution of type 2 diabetes: Impact on glucose tolerance and β -cell function. *J Clin Endocrinol Metab* 89: 2846–2851, 2004.
119. Zhou W, Wang ZL, Kaduce TL, Spector AA, and Lee HC. Impaired arachidonic acid-mediated dilation of small mesenteric arteries in Zucker diabetic fatty rats. *Am J Physiol Heart Circ Physiol* 288: H2210–2218, 2005.

Address reprint requests to:

Dr. David Busija

Department of Physiology and Pharmacology

Wake Forest University Health Sciences

Medical Center Boulevard

Winston-Salem, NC 27157

E-mail: dbusija@wfubmc.edu

Date of first submission to ARS Central, December 28, 2005;
date of acceptance, January 22, 2006.

This article has been cited by:

1. Mona F. Mahmoud, Mohamed El-Nagar, Hany M. El-Bassossy. 2012. Anti-inflammatory effect of atorvastatin on vascular reactivity and insulin resistance in fructose fed rats. *Archives of Pharmacal Research* **35**:1, 155-162. [[CrossRef](#)]
2. Hany M. El-Bassossy, Ahmed Fahmy, Dina Badawy. 2011. Cinnamaldehyde protects from the hypertension associated with diabetes. *Food and Chemical Toxicology* . [[CrossRef](#)]
3. Hany M. El-Bassossy, Mohamed A. El-Moselhy, Mona F. Mahmoud. 2011. Pentoxifylline alleviates vascular impairment in insulin resistance via TNF- α inhibition. *Naunyn-Schmiedeberg's Archives of Pharmacology* . [[CrossRef](#)]
4. Zachary C. Berwick, Gregory M. Dick, Johnathan D. Tune. 2011. Heart of the matter: Coronary dysfunction in metabolic syndrome. *Journal of Molecular and Cellular Cardiology* . [[CrossRef](#)]
5. Eric P. Davidson, Lawrence J. Coppey, Brian Dake, Mark A. Yorek. 2011. Effect of Treatment of Sprague Dawley Rats with AVE7688, Enalapril, or Candoxatril on Diet-Induced Obesity. *Journal of Obesity* **2011**, 1-9. [[CrossRef](#)]
6. Richard M. Cubbon, Matthew B. Kahn, Stephen B. Wheatcroft. 2009. Effects of insulin resistance on endothelial progenitor cells and vascular repair. *Clinical Science* **117**:5, 173-190. [[CrossRef](#)]